OPEN ACCESS

Advising a cardiac disease gene positive yet phenotype negative or borderline abnormal athlete: Is sporting disqualification really necessary?

Pascale Richard,[1,2,3] Isabelle Denjoy,[2,4] Véronique Fressart,[1,3] Mathew G. Wilson,[5] François Carré,[6] Philippe Charron[2,3]

[1]AP-HP, Functional Unit of Cardiogenetics and Myogenetics, department of Biochemistry, Pitié-Salpêtrière Hospital, Paris, France
[2]AP-HP, National referral centre for inherited cardiac diseases, Pitié-Salpêtrière Hospital, Paris, France
[3]UPMC – Paris 6 University, INSERM UMRS-956, Paris, France
[4]Department of Cardiology, AP-HP, Bichat Hospital, Paris, France
[5]Department of Sports Medicine, ASPETAR, Qatar Orthopaedic and Sport Medicine Hospital, Doha, Qatar
[6]Pontchaillou Hospital – Rennes 1 University, fonctionnal explorations Unit, Inserm U 1099, 35033 Rennes cedex, France

Correspondence to
Dr Philippe Charron, Centre de référence pour les maladies cardiaques héréditaires, Hôpital Pitié-Salpêtrière, 47 Bd de l'Hôpital, 75013 Paris, France; philippe.charron@psl.aphp.fr

ABSTRACT

The sudden cardiac death (SCD) of an athlete is a rare and tragic event, often caused by a number of inherited heart muscle disorders, namely the cardiomyopathies and primary arrhythmia syndromes (also known as cardiac ion channelopathies). Recent advances in the understanding of the molecular genetics of these heritable cardiovascular diseases present new challenges for clinicians who manage athletes with these types of heart muscle conditions. Unfortunately, the clinical heterogeneity of many of these SCD diseases are also matched by the genotypic heterogeneity associated with the pathogenesis of the disease. A particularly challenging situation arises when the sports physician and attending cardiologist are presented with an athlete who demonstrates a familial context of inherited cardiac disease or presents mild cardiac abnormalities suggestive of inherited cardiac disease. Alongside the complete cardiac evaluation, genetic testing may be proposed as an additional diagnostic tool in this clinical conundrum. However, debate still remains on how extensive the screening should be, in particular the use and interpretation of genetic testing in that setting. The aim of this review is to examine the role of genetic testing within the diagnostic algorithm of preparticipation screening of athletes. This will be achieved by providing the sports medicine physician with simple current cardiac genetic knowledge for the main inherited cardiac conditions known to cause SCD. Furthermore, it will examine current knowledge for the role of genetic testing upon the prediction of SCD, concluding with its impact on the sport eligibility and disqualification conundrum using case examples from our genetic testing laboratory.

INTRODUCTION

The appropriate diagnosis of an inherited heart disease has become a major issue in preparticipation screening of athletes, as these diseases represent the main causes of sudden cardiac death (SCD) in athletes under the age of 35 years.[1–4] A diagnostic conundrum occurs when the physician is presented with an athlete who demonstrates a familial context of inherited cardiac disease or presents mild cardiac abnormalities suggestive of inherited cardiac disease. Alongside the complete cardiac evaluation, genetic testing may be proposed as an additional diagnostic tool in this challenging situation. Indeed, given the therapeutic strategies now available to prevent sudden death, such as the implantable cardioverter-defibrillator, the necessity for early identification of certain cardiomyopathies or primary arrhythmia syndromes in competitive athletes has become magnified. The aim of this review is to examine the role of genetic testing within the diagnostic algorithm of preparticipation screening of athletes. This will be achieved by providing the sports medicine physician with current cardiac genetic knowledge for the main inherited cardiac conditions known to cause SCD. Furthermore, it will examine current knowledge for the role of genetic testing upon the prediction of SCD, concluding with its impact upon the sport eligibility and the disqualification conundrum.
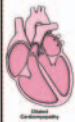
MOLECULAR BASIS OF INHERITED CARDIAC DISEASES AT RISK OF SUDDEN DEATH IN COMPETITIVE ATHLETES

Adrenergic stress during competitive sport is a commonly accepted trigger for arrhythmia and SCD, in the presence of underlying inherited cardiac disease such as cardiomyopathy (diseases with structurally and functionally abnormal myocardium in the absence of common condition sufficient to cause it),[5] primary arrhythmia syndromes (pure electrical diseases mostly due to ion channel dysfunction)[6] or vascular disease (such as Marfan syndrome).[7] In an examination of 1866 young athletic SCD's, the most common cause was hypertrophic cardiomyopathy (HCM) (36%), but other inherited diseases were also frequently observed; namely 'possible' HCM (17%), arrhythmogenic right ventricular cardiomyopathy (ARVC) (4%), long QT syndrome (LQTS) (4%), aortic rupture (3%) and dilated cardiomyopathy (DCM) (2%).[3] A common theme across recent histopathological studies is the under-reporting in the frequency of primary arrhythmia syndromes, as this diagnosis is usually missed by necropsy. For the cardiac geneticists, however, ion channel diseases can be identified via postmortem genetic analyses in up to 10–30% of cases who often demonstrate normal conventional necropsy.[8–10]

INHERITED CARDIAC DISEASE: GENES AND INHERITANCE

The main genes involved in inherited cardiac disorders are indicated in figure 1. The mode of inheritance for most of these diseases is autosomal dominant, but in particular phenotypes such as syndromic diseases or paediatric patients with cardiomyopathies, recessive, X-linked or mitochondrial modes of inheritance may also be observed (figure 1A, bottom). The familial aggregation is not always recognised, making it difficult to conclude about a familial/genetic disease. Sporadic

A

Phenotypes	Protein	Gene symbol	Frequency in mutated patients	Ref.
Hypertrophic Cardiomyopathy	Beta Myosin heavy chain	MYH7	25-35%	16, 63
	Myosin binding protein C, cardiac	MYBPC3	30-50%	16, 63
	Cardiac troponin T	TNNT2	5-7%	16, 63
	Cardiac troponin I	TNNI3	3-5%	16, 63
	Myosin, light chain 2, regulatory	MYL2	~3-5%	16
	Myosin, light chain 3, essential	MYL3	~2%	16
	Alpha Tropomyosin, Alpha Myosin heavy chain, Titin, Cardiac Actin, Telethonin, Myozenin	TPM1, MYH6, TTN, ACTC1, TCAP, MYOZ2	rare	
Dilated Cardiomyopathy	Lamin A/C	LMNA	5- 10%	27-63
	Beta Myosin heavy chain (β MHC)	MYH7	~7%	27
	Cardiac troponin T (cTNT)	TNNT2	~4%	27
	Myosin binding protein C, cardiac (cMyBPC3)	MYBPC3	~1%	27
	Sodium channel, type V, α subunit	SCN5A	5-10%	63
	Cardiac troponin I (cTNI)	TNNI3	~1%	27
	Myosin, light chain 2, regulatory	MYL2	~1%	
	Myosin, light chain 3, essential	MYL3	~1%	
	Desmin	DES	~3%	
	BCL2-associated athanogene 3	BAG-3	~3%	
	Alpha Tropomyosin, Titin, Cardiac Actin, Telethonin, Myozenin,	TPM1, TTN, ACTC1, TCAP, MYOZ2	~2%	
Arrhythmogenic right ventricular cardiomyopathy (ARVC)	Plakophilin 2	PKP2	11-43%	20-59
	Desmoglein 2	DSG2	12-40%	20-59
	Desmoplakin	DSP	6-16%	20-59
	Desmocollin 2	DSC2	rare	20-59
	Junction plakoglobin	JUP	rare	20-59
	Ryanodine receptor 2	RYR2	rare	20-59
	Transmembrane protein 43	TMEM43	rare	20-59
Syndromic Cardiomyopathy	**Protein**	**Gene symbol**	**Inheritance**	
HCM+WPW (Wolff Parkinson White)	Protein kinase, AMP-activated, γ 2	PRKAG2	Autosomal Dominant	
Danon disease	Lysosomal-associated membrane protein 2	LAMP2	X linked	
Pompe disease	alpha Glucosidase	GAA	Recessive	
Fabry disease	alpha Galactosidase	GLA	X Linked	
Friedreich syndrome	Frataxin	FXN	Recessive	
Marfan Syndrome	Fibrillin 1	FBN1	Dominant	

Figure 1A Main phenotypes and genes responsible for cardiovascular inherited diseases with their respective frequency. (A) Cardiomyopathies and (B) rhythm disturbances. This figure is only reproduced in colour in the online version.

cases should be considered as part of genetic disease (usually with autosomal dominant inheritance), especially for HCM and ARVC, either because of a de novo mutation (mutation not transmitted from a parent) or because of incomplete penetrance in the parent (mutation without cardiac expression).[11–13]

HCM is characterised by the presence of increased ventricular wall thickness or mass in the absence of loading conditions (hypertension, valve disease, etc) sufficient to cause the observed abnormality[5] with a disease prevalence of 1/500.[14] The usual mode of inheritance is autosomic dominant, characterised by genetic heterogeneity, with more than 900 different mutations described in more than 13 genes[15–18] (Sarcomere Protein Gene Mutation Database http://cardiogenomics.med. harvard.edu). These genes encode the proteins that form various components of the contractile apparatus of the sarcomere. The global efficiency of mutation screening in index cases is of 30 to 65%. A study of nearly 200 independent HCM patients reported that 42% of genotype-positive patients had mutations in myosin-binding protein C (MYBPC3), 40% in β-myosin heavy chain (MYH7), 6% in troponin T (TNNT2), 6% in troponin I (TNNI3) and 4% in regulatory myosin light chain (MYL2)(figure 1A).[16]
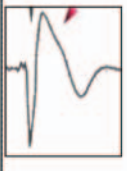
B

Phenotypes	Protein	Gene symbol	Freq.	Ref
Long QT Syndrome (LQTS)	Potassium voltage- gated channel, member 1 (KVLQT1)	*KCNQ1*	58%	35, 63
	Potassium voltage-gated channel, member 2 (KCNH2)	*KCNH2*	35%	35, 63
	Potassium voltage gated channel, member 1	*KCNE1*	1%	35, 63
	Potassium voltage-gated channel, member 2	*KCNE2*	1%	35
	Sodium channel, voltage gated, type V, α peptide	*SCN5A*	5%	35
	Potassium channel, subfamily J, member 2 (Kir2.1)	*KCNJ2*	rare	
	Calcium channel, voltage-dependent, alpha 1C	*CACNA1C*	rare	
	Caveolin 3	*Cav3*	rare	
	Sodium channel, voltage-gated, type IV, α subunit	*SCN4A*	rare	
	A-kinase anchor protein 9	*AKAP9*	rare	
	Syntrophin	*SNTA1*	rare	
	Potassium channel, subfamily J, member 5 (Kir3.4)	*KCNJ5*	rare	
Brugada Syndrome (BS)	Sodium channel, voltage gated, type V, α subunit	*SCN5A*	15%	63
	Glycerol-3-phosphate déshydrogenase 1-like	*GPD1L*	rare	
	Calcium channel, voltage-dependent, α1C subunit (Cav1.2)	*CACNA1C*	rare	
	Calcium channel, voltage-dependent, β 2 subunit	*CACNB2*	rare	
	Sodium channel, type I, beta subunit (Navβ1)	*SCN1B*	rare	
	Potassium voltage-gated channel, member 3	*KCNE3*	rare	
	Sodium channel, voltage-gated, type III, β subunit	*SCN3B*	rare	
	Hyperpolarization activated cyclic nucleotide-gated potassium channel 4	*HCN4*	rare	
	Potassium inwardly-rectifying channel, subfamily J, member 8	*KCNJ8*	rare	
	Calcium channel, alpha 2/delta subunit 1 (Cav2δ)1	*CACNA2D1*	rare	
	Voltage-gated potassium (Kv) channel	*KCNE5*	rare	
	Multicopy suppressor Of Gsp1	*MOG1*	rare	
	Potassium voltage-gated channel, Shal-related subfamily, member 3 (Kv4.3)	*KCND3*	rare	
Short QT Syndrome (SQTS)	Potassium inwardly-rectifying channel, subfamily J, member 2	KCNJ2	rare	63
	Potassium voltage- gated channel, member 1	KCNQ1	rare	63
	Potassium voltage-gated channel, member 2	KCNH2	rare	63
	Sodium channel, type V, αpolypeptide	SCN5A	rare	63
Cathecholaminergic polymorphic ventricular tachycardia (CPVT)	Ryanodine receptor 2	RYR2	60%	63
	Cardiac Calsequestrin 2	CASQ2	rare	

Figure 1B continued.

ARVC has an estimated prevalence of 1/2000–1/5000 in the general population and is histologically characterised by the replacement of myocytes within the right ventricular myocardium by adipose and fibrous tissue.[19][20] The disease is transmitted as an autosomal-dominant trait, with five of the responsible genes encoding the major components of the cardiac desmosome (figure 1A). According to the different number of published studies, the global efficiency of mutation detection is 30–70% in index cases. Data from our laboratory of 135 independent French patients indicate that plakophilin-2 (*PKP2*) is the predominant gene mutation (31% of patients), followed by desmoglein-2 (*DSG2*, 10%), desmoplakin (*DSP*,

4.5%), desmocollin-2 (*DSC2*, 1.5%) and junction plakoglobin (*JUP*, 0%).[21]

DCM is defined by left ventricular (LV) dilatation and systolic dysfunction in the absence of abnormal loading conditions or coronary artery disease, sufficient to cause the impairment and prevalence is estimated to be 1/2500.[5][22] DCM is a sporadic disease in approximately 65% of cases and a familial disease in 35%. Familial forms are characterised by a great genetic heterogeneity (>30 causal genes reported), as well as a huge diversity of related proteins and underlying pathways such as nuclear envelope, cardiac sarcomere, ion channels, transcription factors and dystrophin-associated cytoskeletal complex.[12][23][24]

However, the yield of genetic testing for DCM is relatively low, with a global rate of mutation detection of around 20–30% in index cases, and most mutations related to lamin A/C (*LMNA*, ~5–10%), β-myosin heavy chain (*MYH7*, ~3–7%), cardiac troponin T (*TNNT2*, ~4%), myosin-binding protein C (*MYBPC3*, <1%), sodium channel (*SCN5A*) and α-myosin heavy chain (*MYH6*) (figure 1A).[25–28]

Among the primary arrhythmia syndromes, LQTS is characterised by prolonged QT interval on ECG, syncope and sudden cardiac death due to ventricular tachyarrhythmias.[29 30] The prevalence is estimated to be 1 in 2500 in the general population[31 32] but two studies reported a high prevalence of LQTS founder mutations in the Finnish (0.4%) and Norwegian (1%) populations.[33 34] LQTS is usually of genetic origin, with autosomal-dominant inheritance and an efficiency of mutation identification of 40–60% in index cases.[29 35] Three common forms of LQTS represent the majority of cases; LQT1, a slow outward delayed rectifier potassium current abnormality encoded by mutations of *KCNQ1* gene; LQT2, a rapid component outward delayed rectifier potassium current abnormality encoded by mutation of *KCNH2* gene and; LQT3, a gain-of-function sodium channel abnormality encoded by mutation of *SCN5A* gene. Mutations in *KCNQ1* represent 50% of genotyped LQTS, *KCNH2* 40% and *SCN5A* accounts for less than 5% of LQTS patients.[36] However, it should be noted that some rare mutations have been identified in the auxiliary subunits of ion channels (figure 1B).[6 37–39]

There are several other inherited cardiac diseases that can cause sudden death that the genetic cardiologist may face within the athletic community. Catecholaminergic polymorphic ventricular tachycardia (CPVT) is a rare arrhythmogenic disorder (prevalence 1/10 000) characterised by adrenergic-induced bidirectional and polymorphic ventricular tachycardia.[30, 32] Responsible genes for CPVT encode the cardiac ryanodine receptor (*RYR2*) and the cardiac calsequestrin 2 (*CASQ2*; figure 1B). Despite rarely a cause of SCD in athletes, yet worthy of mention in this review are the conditions of Brugada syndrome or short QT syndrome. The global rate of mutation identification in index cases is 65% in CPVT and 20–30% in Brugada syndrome.[40]

PENETRANCE OF MUTATIONS AND NATURAL HISTORY OF THE DISEASES
Penetrance is defined by the proportion of mutation carriers with an obvious clinical expression, such as an abnormal ECG and/or echocardiogram. However, when considering the lifetime cumulative risk of expressing the phenotype, penetrance may be complete (100% risk of developing the disease with age) or incomplete (<100% risk of developing the phenotype). Mutations responsible for inherited cardiac diseases exhibit a variable penetrance, typically increasing with age. The onset of the cardiac expression (phenotype) is therefore usually delayed with a strictly normal cardiac examination for many years, and may explain why some athletes may perform extraordinary physical achievements for many years without any detection of inherited cardiac disease. The general natural history of these diseases is normally characterised by three important progressions; (1) a silent stage without personal symptoms, normal cardiac examinations and a normal ECG/echocardiogram (NB: the mutation is already detectable at this stage), (2) a second stage without symptoms but with detectable cardiac expression of the disease on ECG or echocardiogram (frequently delayed after 20 years of age for the majority of cardiomyopathies) and (3) a third stage with symptoms, obvious cardiac expression of the disease or sometimes the occurrence of unexpected SCD.

When considering the lifetime risk of developing the disease, penetrance may be nearly 'complete' after 50–60 years of age, in most cases in HCM or DCM (penetrance: ~95%),[11 41] whereas for other mutations or diseases such as ARVC, LQT and Brugada syndromes, incomplete penetrance is commonly observed.[6 42] Furthermore, the age for the phenotypic expression onset is also different according to the disease type, with frequent cardiac expression in young children and even neonates for LQTS but infrequent expression before 10 years of age for HCM or ARVC. Penetrance is also influenced by gender, as the expression of disease is more frequent with earlier onset in males for HCM, DCM, ARVC and Brugada syndrome, whereas LQTS cardiac expression is more frequent in females.[6 11 43]

VARIABLE EXPRESSIVITY AND GENOTYPE–PHENOTYPE CORRELATIONS
Patients with an inherited cardiac disease usually present a variable severity in phenotypic expression, called expressivity, with dramatic differences in the age at onset, degree of symptoms, severity of ECG/echocardiography features, risk of complications and response to medical treatment. This variability is observed in patients sharing the same genetic mutation, both at interfamilial and intrafamilial levels, suggesting that epigenetic factors as well as environmental factors may act on the severity of the phenotype.[12]

However, the variability in the disease severity is accentuated between families by a great genetic heterogeneity with multiple genes, multiples mutations within the genes, various pathogenic mechanisms of mutations that could be involved in a given patient. The precise underlying gene and mutation may contribute to the variable expressivity of the phenotype,[6 44 45] what is known as phenotype–genotype correlations and may have important consequences for genetic counselling and medical management of the athlete. Such correlations have been observed although they still require to be confirmed by the study of large populations and a prospective follow-up.

In HCM patients, genotype–phenotype correlations have shown that mutations in troponin T (*TNNT2*) usually cause mild hypertrophy that can escape clinical detection, but are associated with a high risk of premature SCD[15 46 47] whereas mutations in myosin-binding protein C (*MYBPC3*) have been associated with delayed onset of HCM and significantly lower risk of SCD.[48 49] In DCM patients, high mortality related to both heart failure and premature SCD have been described in patients with *LMNA* mutations.[50] In ARVC patients, a high risk of left ventricle dysfunction was associated with desmoplakin (DSP) or desmoglein-2 (*DSG2*) mutations.[21 51] In LQTS patients, cardiac events are often determined by the underlying genotype. In LQT1 phenotype, involving mutations in *KCNQ1*, cardiac events are mostly triggered by adrenergic stimulation, typically exercise. In LQT2 phenotype, involving mutations of *KCNH2*, cardiac events are triggered by emotion and in LQT3 patients with mutations in *SCN5A* cardiac events are triggered by sleep/rest. Efficacy of β-blocker treatment is also variable in LQTS patients according to the underlying gene: very efficient to reduce syncope and SCD in LQT1, whereas less efficient in patients with *SCN5A* mutations.[6 52–54]

Not only the gene but also the precise mutation within a given gene might be important. For example, in LQT1 patients, the higher risk of SCD concerns patients with mutations in the transmembrane domains of the protein encoding for the pore of the channel[55 56] and in DCM patients, a higher risk of SCD was associated with the *LMNA* mutations that led to truncated proteins (mutations disrupting the reading frame during translation).[50]

Recent molecular findings also shed light on the intrafamilial variability of these diseases, especially in HCM and ARVC. Even in the context of familial forms and a dominant inheritance, a subset of the patients (3–10%) may carry more than one heterozygous mutation, so-called multiple mutations (either in the same gene or in another gene). Interestingly, these patients exhibit a more severe phenotype and a more frequent risk of SCD.[21] [57–59] Additional factors may also modulate the phenotype and contribute to the intrafamilial variability, such as common genetic polymorphisms.[60] [61] or acquired/environmental factors (such as myocarditis), but they are still poorly understood and further research is required in this specialised area.

GENETIC TESTING IN PRACTICE

Genetic testing was developed in some authorised medical laboratories for the purpose of routine use in order to improve both genetic counselling and the medical management of patients and relatives with inherited cardiac pathologies,[11] [26] [62] [63] and these basic principles are applicable to all populations including high-level athletes.

The molecular testing begins with a blood sampling of an index patient with an obvious form of a disease (with a certain or possible genetic origin), for which genes of interest are tested for mutation by sequencing (determination of the primary sequence of genes). For most cardiac diseases, the analysis concerns several genes and all the coding sequences of each of these genes. The process can therefore be quite long-winded and results for the index patient may often take several months. In HCM and ARVC, genes are tested according to their frequency but in patients with a family history of SCD, *TNNT2* can be first tested. A suggestive phenotype can orientate the screening, as in DCM with conduction defect where the *LMNA* gene is the first tested and in LQTS patients where the ECG guides the testing. The usual efficiency of a conventional mutation screening in a medical laboratory may therefore vary from quite high (50–70% in HCM, LQTS or CPVT and ~50% in ARVC) to quite low (20–30% in DCM or Brugada syndrome). The efficiency of the mutation screening depends not only on the complexity of the underlying molecular data, that is, the number of genes to screen, the large size of these genes and the sensitivity of the methods, but also on a careful characterisation of the phenotype and the possibility of phenocopy or environmentally induced phenotype mimicking a genetically determined trait. For example, a patient with concentric HCM associated with a Wolf Parkinson White syndrome will be tested first on *PRKAG2* gene before sarcomeric genes.

In a few cardiac diseases with a particular phenotype, the analysis can be focused on one or few more predominant mutations within a given gene as in haemochromatosis due to mutations in the hereditary haemochromatosis protein encoded by the HFE gene, familial amyloidosis with mutations in transthyretin or Marfan syndrome with mutations in fibrillin-1 gene. The result is then available in few days or weeks with a very high efficiency.

Conversely, when the causal mutation is identified in the index-patient of the family, it is easy (one PCR/sequence reaction), quick (within few days) and efficient (100% result: the familial mutation is present or absent) to determine the genetic status of any relative within this family.

The recent evolution of sequencing technologies and their use in the very near future for routine diagnosis will dramatically change the practice and interpretation of genetic testing. Next Generation Sequencing (NGS) is used to designate new methods of sequencing that provide massive high throughput sequencing in a short time.[64] [65] NGS can be used for the simultaneous analysis of numerous target disease or candidate genes for a given disease. In inherited cardiomyopathies and rhythm disturbances, 50–70 disease genes could be analysed simultaneously. This approach will undoubtedly revolutionise the diagnostic approach of genetic medicine but it also poses many new challenges, not least the analysis of massive amounts of data and the interpretation of the many tens or thousands genetic variants that will be identified in individual patients or athletes.

GENETIC TESTING AND THE INTERPRETATION OF RESULTS IN ATHLETES

In the case of an athlete with a suspected cardiomyopathy or ion channelopathy, there are two different situations for the recommendation of mutation analysis to be performed (figure 2).
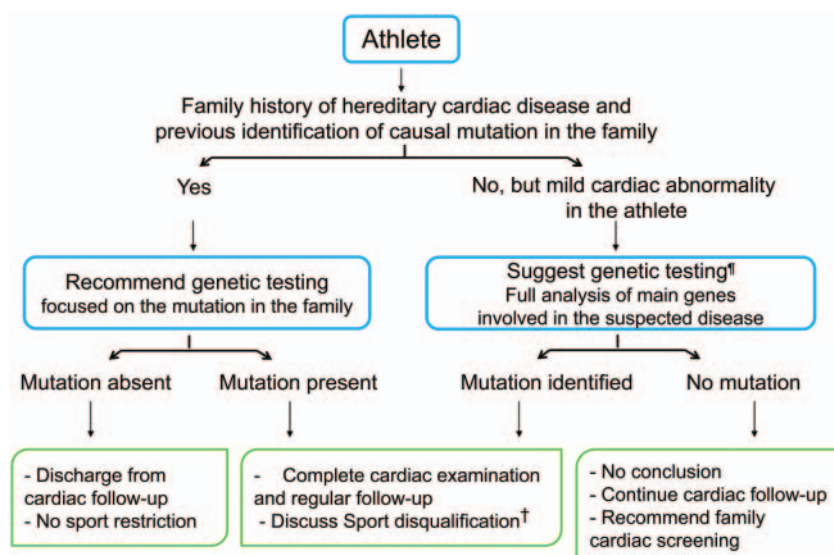
Figure 2 Strategy for genetic testing in atheletes with a familial context of inherited cardiac disease or with mild abnormalities. ¶Decision by specialized multidisciplinary teams †Case by case approach according to the disease, cardiac features, gene, mutation, specific sport that is practiced. This figure is only reproduced in colour in the online version.

The first situation is when there is a family history of cardiac hereditary disease and the mutation is identified in the athlete's family. Genetic testing can be proposed to the athlete (asymptomatic with a normal or borderline cardiac examination) in order to search for the presence or absence of the specific mutation previously identified in their family. The process leads to a rapid and appropriate answer whereby (1) the presence of the mutation in the athlete is identified, leading to complete cardiac examination, and the discussion on specific preventive therapy or life-style modifications, together with effective management and regular medical follow-up or (2) the identified mutation in the family is absent within the athlete. Thus, the athlete can be discharged without the need for further cardiac examination; only the basic risk of the disease prevalence in the general population remains. *The second situation* is when there is no family history of cardiac hereditary disease, but the athlete may present borderline cardiac abnormalities on ECG or echocardiography and/or symptoms, however not sufficient per se to confirm a diagnosis of a cardiomyopathy or primary cardiac arrhythmia. In such cases, where there is adequate evidence to consider the hypothesis of an inherited disease, genetic testing can be proposed directly to the athlete.

If a mutation is identified, then the diagnostic information is crucial for the athlete and their family, leading to appropriate cardiac examination and long-term management. The interpretation of the genetic variant is, however, a very important step that requires several criteria to be studied[66] including (1) the absence from an ethnically matched control population of at least 400 chromosomes and from web access polymorphisms data bases; (2) the nature of the mutation, where non-sense mutations as insertions, deletions and stop codons are usually pathogenic verses substitution of amino acids that may be questionable and (3) in silico modelisation of the consequences (location of the mutated residue in a conserved amino acid sequence, predicted secondary structure regions from published reports and the databank such as UniProtKB/Swiss-Prot). If no mutation is identified in this second situation, no conclusive diagnosis can be drawn since a mutation is not always identified in patients with obvious familial disease. Finally, if only a genetic variant of unknown significance is identified in this second situation, then no conclusion can be drawn. The interpretation of the result in this second scenario is therefore very important and it should be borne in mind that only a positive result (identification of a pathogenic mutation) is meaningful.

RISK OF CARDIAC DEATH AT THE EARLY STAGE OF INHERITED CARDIAC DISEASES

The risk of cardiac complications in patients with an obvious form of inherited cardiac disease is well known, as well as the deleterious role of intensive or competitive sport practice through the occurrence of ventricular arrhythmia. For athletes, specific therapeutic management often includes high-intensity sporting restriction, as underlined by international consensus recommendations.[67–70] In contrast, however, the risk of a cardiac event at an earlier stage, in athletes carrying a causal mutation but without obvious cardiac expression of the disease, is poorly understood.

Hypertrophic cardiomyopathy

A limited number of sudden deaths have been described in asymptomatic patients without overt hypertrophy but carrying sarcomeric gene mutation.[71 72] McKenna et al[73] described two families with five premature sudden deaths (from 17 to 44 years) in patients without macroscopic hypertrophy, but with widespread myocardial disarray on histological examination, and ECG repolarisation abnormalities in alive relatives in a family. Several years later, the p.Arg94Leu mutation in the troponin T (*TNNT2*) gene was identified in this family in two relatives with abnormal ECG and in one of the deceased patients (postmortem analysis).[71] Another report described two patients resuscitated from a cardiac arrest caused by ventricular fibrillation in which a clinical diagnosis of HCM could not be made at the time of the event but after several years due to the evolution of the phenotype, and because of the diagnosis of HCM in a family member. A mutation in the β-myosin heavy chain (*MYH7*) gene was eventually identified in each patient (p.Leu517Arg and p.Arg858Leu, respectively).[72] Comprehensive risk stratification for SCD in sarcomeric mutation carriers without clinical HCM expression is poorly understood at the present time. The most recent and largest study was performed in 339 mutation carriers without clinical HCM.[74] One risk factor for SCD was observed in 30% of the mutation carriers (including 4% with non-sustained VT and 14% with abnormal blood pressure response during exercise) and ≥2 risk factors were observed in only approximately 3% of mutation carriers. However, a short follow-up (3.5 years) study of sarcomeric mutation carriers without clinical HCM expression revealed only two SCD (0.13% per person-year) and suggests that for this population with 82% of patients with an *MYBPC3* mutation, a good prognosis can be observed.[74] On the contrary, in a study of 75 *TNNT2* mutation carriers including 34 without clinical HCM at baseline, the rate of major cardiac events was 0.93% person-year suggesting the role of the mutated gene in the risk stratification.[75] Although it is unclear at present as to whether risk factors for SCD are also of prognosis significance for HCM mutation carriers without manifested disease; thus, the findings suggest careful follow-up of the mutation carrier.

Arrhythmogenic right ventricular cardiomyopathy

ARVC is characterised by a high occurrence of SCD, even as a first clinical manifestation as suggested by the relatively frequent identification of ARVC at necropsy after unexpected SCD.[76 77] Bauce et al[78] described the case of a 13-year-old *DSP* mutation carrier, considered as clinically unaffected during familial screening, who died suddenly 2 years later (ECG performed 8 months before death for sports eligibility demonstrated only incomplete right bundle branch block). In a large study of 12 500 trained athletes in Italy,[68] 81 were found with abnormal negative T waves on ECG and no apparent cardiac disease. After a follow-up of 12±5 years, five athletes ultimately proved to have a cardiomyopathy including one who died suddenly at 24 years from clinically undetected ARVC, together with four living patients who eventually developed HCM (three cases) or DCM (one case).

Dilated cardiomyopathy

Premature SCD has been described in early stage of DCM, specifically related to *LMNA* mutations that are associated with early conduction defects and/or ventricular arrhythmia.[50 79] In a study of 19 *LMNA* mutation carriers with normal LV ejection fractions but with significant conduction defects requiring pace maker, a defibrillator was also systematically implanted and a high rate of appropriate implantable cardioverter defibrillator (ICD) therapy was observed (42% at 34±21 months), indicating a high prevalence of ventricular arrhythmia and thus, risk of sudden death before the onset of cardiac failure.[80] Moreover, the recent case

study report of the SCD of a 35-year-old woman with a *LMNA* mutation (c.908-909delCT), who demonstrated normal LV function and without significant conduction disease,[81] suggests the possibility that early and severe ventricular arrhythmia may be the only phenotypic expression of the disease.

Long QT syndrome

A recent study observed that the cumulative probability of aborted cardiac arrest or SCD in LQTS mutation carriers with normal-range QTc intervals (<440 ms) was significantly lower than in those with prolonged QTc intervals (4% vs 15%, p<0.001); but was however significantly higher than in those unaffected family members without mutation (4% vs 0.4%, p<0.001).[55] Importantly, the authors identified subgroups of genotype-positive yet phenotype-negative relatives who were at significant risk of SCD (ie, LQT1 and LQT3 with transmembrane-missense mutations). This specific subgroup of mutation carriers with normal QTc interval might therefore benefit from early β-blockers intervention, in addition to sport restriction and avoidance of QT-prolonging medications.

Apart from the acute risk of sudden death at the early stage of an inherited cardiac disease possibly due to the interaction between an arrhythmogenic substrate, adrenergic stress and/or a premature ventricular beat, the potential role of regular intensive sport activity on the development of myocardial dysfunction (diastolic or systolic) has been questioned in the context of cardiomyopathies. Although still speculative, in human[82] and animal models, some have suggested a potential deleterious effect. For example, in a mutated ARVC mouse model, endurance training accelerated the development of right ventricular dysfunction (and arrhythmia inducibility) in heterozygous plakoglobin-deficient mice, suggesting that endurance training could accelerate disease occurrence or progression.[83]

MANAGEMENT OF ATHLETES WITH A MUTATION BUT A NORMAL OR BORDERLINE CARDIAC EXAMINATION

To evaluate the risk of SCD associated with competitive sports in the presence of a cardiomyopathy or primary arrhythmia syndrome is a dilemma for the attending cardiologist. This has led to several international recommendations for the preparticipation screening in competitive sports (focused on the identification of a cardiac disease not recognised until now),[84 85] as well as recommendations for participations of patients (with a recognised cardiac disease) for competitive sports or leisure time physical activities.[67 68 86–89] The practical management of athletes however remains a challenging issue, as little longitudinal data exist supporting the beneficial role of sporting disqualification upon death rates of athletes identified with an inherited cardiac disease, outside of the Italian experience. Because of this death of data, sporting disqualification should also be viewed as the last intervention available in the cardiologist's armoury. It can be anticipated that with future improvement in the understanding of such diseases and their potential risk of adverse cardiac events associated with intensive sport, the objective should and will be to reduce the number of unnecessary disqualifications and to modulate rather than stop the sports activity of the athlete in question.

It is important to realise that the decision for sporting disqualification is based upon the stage of the disease when it is identified within the athlete. This is due to the fact that the natural history of inherited cardiac diseases is often characterised by a dynamic process and a continuum from (1) an asymptomatic mutation carrying athlete in the absence of cardiac expression or who demonstrates mild-to-borderline abnormalities,

(2) an asymptomatic athlete with obvious cardiac expression with or without specific SCD risk factors such as non-sustained ventricular tachycardia in HCM or extremely prolonged QTc interval in LQTS patient and finally (3) an athlete with obvious cardiac expression who presents the occurrence of symptoms and/or cardiac complications. Although the risk of SCD is not always strictly related to the stage of cardiac expression, there is a general consensus on the disqualification of competitive sport in athletes with obvious cardiac expression.[68 86 87 89] These recommendations are supported by the reduced number of sudden deaths in Italy since the development of systematic preparticipation screening in athletes.[90]

The management of asymptomatic athletes who carry a mutation but without any cardiac expression or with only borderline abnormalities, is a particularly challenging issue without consensus.[68] The risk of sudden death at this early stage is probably low but not negligible, although available data are limited. In the context of such 'genotype-positive–phenotype-negative' athletes, there are significant differences between European and North American policy. Study Groups of the European Society of Cardiology recommend[86] the disqualification of all mutation carriers (especially within HCM or Marfan athletes), whereas the North Americans (Task Force 4) authorise full competitive sport (except for swimming in individuals with LQT 1 mutation).[86, 87]

In our institutions, the management of genotype-positive–phenotype-negative individuals and the subsequent discussion regarding sporting restriction or clearance is based on a case-by-case approach taking into account several factors: (1) the type of gene and mutation identified in the athlete, as some mutations have been acknowledged as being at either a high or low risk for an adverse cardiac events; (2) the familial history and number of premature cardiac deaths as several sudden deaths before 50 years of age in a family represent a known risk factor for HCM and can be considered important in the discussion of sporting restriction with the athlete. Unfortunately, for most primary arrhythmia syndromes, these scientific evidence is lacking; (3) the type of sporting activity undertaken, as highly dynamic sports have been linked with SCD particularly soccer and basketball for HCM and swimming for LQT and finally (4) the presence in the athlete of personal SCD risk factors after prognostic stratification such as non-sustained ventricular tachycardia in HCM mutation carriers.

The following two cases examples from our institutions illustrate the clinical value of genetic testing in the diagnosis of the condition, thus helping in the discussion of sporting restriction or medical clearance for the athlete. One case was brought in as an athlete with symptoms, whereas the second case is 'predictive' in the context of a recognised inherited disease in the athlete's family.

Case 1

A 38-year-old male swimmer (training ~6 h/week) was admitted in our cardiology intensive care unit following multiple episodes of faintness that occurred during swimming. Electrocardiogram at admission demonstrated bradycardia (35/min) with high-degree atrio-ventricular block. Family history questioning revealed a paternal aunt that died suddenly at age 65 years of age. Echocardiography revealed HCM with a basal diastolic septal thickness of 18 mm, a LV diastolic diameter of 42 mm, and a normal LV ejection fraction (60%). Consequently, a pacemaker was implanted the following day. Genetic testing was performed and the analysis was negative for some genes including the *LMNA* gene. *PRKAG2* sequencing revealed a

heterozygous missense disease causing mutation (p.Ser548Pro), responsible for both conduction defect and the HCM expression. This result allows for effective family management and screening, thus helping to avoid potential SCD in relatives.

Case 2

A 14-year-old asymptomatic male soccer player (body surface area 1.59 m^2, training 11 h/week) presented to our department for preparticipation cardiac screening for entry into a professional soccer school, due to family history. The boy's father developed DCM, ICD was implanted and a heterozygous mutation in the *LMNA* gene was identified. The boy's grandfather had a pace maker at 55 years, but died 5 years later. ECG of the boy was unremarkable. Echocardiography exhibited moderate enlarged LV end diastolic diameter (LVEDD 53 mm or 33 mm/m^2) with normal ejection fraction (60%) and wall thickness (7 mm). Exercise test, holter ECG and creatine kinase dosage were normal. In our and others' experience, isolated enlarged LVEDD has been identified in several cohorts of DCM families, as an early predictive marker of relatives at risk of developing latter obvious DCM.[91] After 3 months of detraining, LVEDD was not modified, thus predictive genetic testing was proposed that was accepted following extensive family genetic counselling. Genetic analysis revealed the presence of the *LMNA* mutation in the boy. Owing to the player's early cardiac expression and family history, we proposed that competitive sport was not compatible due to the risk of future occurrence of early ventricular arrhythmia and atrio-ventricular block, in addition to the risk of DCM. Both the boy and the parents agreed on this course of action, with the boy modifying his career choice.

CONCLUSION

Genetic testing is increasingly performed in the clinical arena, leading to the early identification of athletes at risk of developing a potential life-threatening cardiac disease. The management of asymptomatic athletes who carry a mutation but without any cardiac expression, or with only borderline abnormalities, is a particularly challenging issue without international consensus. The risk of sudden death at this early stage is probably low but not negligible. Accordingly, a conservative management is most favourable with education regarding personal symptoms and annual cardiac follow up; with sporting disqualification viewed as a last resort option. We propose that the management of the genotype-positive–phenotype-negative athlete and the subsequent discussion of sporting clearance or restriction is based upon a case-by-case approach taking into account: (1) the type of gene and mutation identified in the athlete, (2) the familial history and number of premature cardiac deaths, (3) the type of sporting activity undertaken and (4) the presence of personal SCD risk factors after prognostic stratification.

Future prospective studies of large cohorts of genotyped relatives are required to improve the knowledge on the risk of SCD, helping to provide optimal management strategies for these athletes. The recent technological advances that enable high-throughput sequencing of multiple genes simultaneously will also revolutionise the diagnostic approach of genomic medicine but in turn will pose many new challenges for the clinician. However, it is anticipated that with the improvements of our understanding of inherited cardiac diseases and their potential risk for adverse cardiac events associated with intensive sport, the sports cardiology community will be able to reduce the number of unnecessary disqualifications and to modulate rather than stop the sporting activity of the athlete in question.

REFERENCES

1. **Corrado D,** Basso C, Rizzoli G, et al. Does sports activity enhance the risk of sudden death in adolescents and young adults? *J Am Coll Cardiol* 2003;**42**:1959–63.
2. **de Noronha SV,** Sharma S, Papadakis M, et al. Aetiology of sudden cardiac death in athletes in the United Kingdom: a pathological study. *Heart* 2009;**95**:1409–14.
3. **Maron BJ,** Doerer JJ, Haas TS, et al. Sudden deaths in young competitive athletes: analysis of 1866 deaths in the United States, 1980–2006. *Circulation* 2009;**119**:1085–92.
4. **Chandra N,** Papadakis M, Sharma S. Preparticipation screening of young competitive athletes for cardiovascular disorders. *Phys Sportsmed* 2010;**38**:54–63.
5. **Elliott P,** Andersson B, Arbustini E, et al. Classification of the cardiomyopathies: a position statement from the European Society of Cardiology Working Group on Myocardial and Pericardial Diseases. *Eur Heart J* 2008;**29**:270–6.
6. **Cerrone M,** Priori SG. Genetics of sudden death: focus on inherited channelopathies. *Eur Heart J* 2011;**32**:2109–18.
7. **Jondeau G,** Michel JB, Boileau C. The translational science of Marfan syndrome. *Heart* 2011;**97**:1206–14.
8. **Tester DJ,** Ackerman MJ. Postmortem long QT syndrome genetic testing for sudden unexplained death in the young. *J Am Coll Cardiol* 2007;**49**:240–6.
9. **Larsen MK,** Nissen PH, Berge KE, et al. Molecular autopsy in young sudden cardiac death victims with suspected cardiomyopathy. *Forensic Sci Int* 2012;**219**:33–8.
10. **Skinner JR,** Crawford J, Smith W, et al. Prospective, population-based long QT molecular autopsy study of postmortem negative sudden death in 1 to 40 year olds. *Heart Rhythm* 2011;**8**:412–19.
11. **Charron P,** Arad M, Arbustini E, et al. Genetic counselling and testing in cardiomyopathies: a position statement of the European Society of Cardiology Working Group on Myocardial and Pericardial Diseases. *Eur Heart J* 2010;**31**:2715–26.
12. **Watkins H,** Ashrafian H, Redwood C. Inherited cardiomyopathies. *N Engl J Med* 2011;**364**:1643–56.
13. **Gandjbakhch E,** Fressart V, Bertaux G, et al. Sporadic arrhythmogenic right ventricular cardiomyopathy/dysplasia due to a de novo mutation. *Europace* 2009;**11**:379–81.
14. **Ommen SR.** Hypertrophic cardiomyopathy. *Curr Probl Cardiol* 2011;**36**:409–53.
15. **Bos JM,** Towbin JA, Ackerman MJ. Diagnostic, prognostic, and therapeutic implications of genetic testing for hypertrophic cardiomyopathy. *J Am Coll Cardiol* 2009;**54**:201–11.
16. **Richard P,** Charron P, Carrier L, et al. Hypertrophic cardiomyopathy: distribution of disease genes, spectrum of mutations, and implications for a molecular diagnosis strategy. *Circulation* 2003;**107**:2227–32.
17. **Marian AJ.** Hypertrophic cardiomyopathy: from genetics to treatment. *Eur J Clin Invest* 2010;**40**:360–9.
18. **Ghosh N,** Haddad H. Recent progress in the genetics of cardiomyopathy and its role in the clinical evaluation of patients with cardiomyopathy. *Curr Opin Cardiol* 2011;**26**:155–64.
19. **Azaouagh A,** Churzidse S, Konorza T, et al. Arrhythmogenic right ventricular cardiomyopathy/dysplasia: a review and update. *Clin Res Cardiol* 2011;**100**:383–94.
20. **Basso C,** Corrado D, Marcus FI, et al. Arrhythmogenic right ventricular cardiomyopathy. *Lancet* 2009;**373**:1289–300.
21. **Fressart V,** Duthoit G, Donal E, et al. Desmosomal gene analysis in arrhythmogenic right ventricular dysplasia/cardiomyopathy: spectrum of mutations and clinical impact in practice. *Europace* 2010;**12**:861–8.
22. **Morimoto S.** Expanded spectrum of gene causing both hypertrophic cardiomyopathy and dilated cardiomyopathy. *Circ Res* 2009;**105**:313–15.
23. **Fatkin D,** Otway R, Richmond Z. Genetics of dilated cardiomyopathy. *Heart Fail Clin* 2010;**6**:129–40.
24. **Villard E,** Perret C, Gary F, et al. A genome-wide association study identifies two loci associated with heart failure due to dilated cardiomyopathy. *Eur Heart J* 2011;**32**:1065–76.
25. **Taylor MR,** Fain PR, Sinagra G, et al. Natural history of dilated cardiomyopathy due to lamin A/C gene mutations. *J Am Coll Cardiol* 2003;**41**:771–80.
26. **Hershberger RE,** Morales A, Siegfried JD. Clinical and genetic issues in dilated cardiomyopathy: a review for genetics professionals. *Genet Med* 2010;**12**:655–67.
27. **Hershberger RE,** Siegfried JD. Update 2011: clinical and genetic issues in familial dilated cardiomyopathy. *J Am Coll Cardiol* 2011;**57**:1641–9.
28. **Lakdawala NK,** Funke BH, Baxter S, et al. Genetic testing for dilated cardiomyopathy in clinical practice. *J Card Fail* 2012;**18**:296–303.
29. **Vincent GM,** Timothy KW, Leppert M, et al. The spectrum of symptoms and QT intervals in carriers of the gene for the long-QT syndrome. *N Engl J Med* 1992;**327**:846–52.

30. **Monteforte N,** Priori SG. The long QT syndrome and catecholaminergic polymorphic ventricular tachycardia. *Pacing Clin Electrophysiol* 2009;**32**(Suppl 2):S52–7.

31. **Goldenberg I,** Moss AJ. Long QT syndrome. *J Am Coll Cardiol* 2008;**51**:2291–300.

32. **Schimpf R,** Veltmann C, Wolpert C, *et al*. Channelopathies: Brugada syndrome, long QT syndrome, short QT syndrome, and CPVT. *Herz* 2009;**34**:281–8.

33. **Berge KE,** Haugaa KH, Früh A, *et al*. Molecular genetic analysis of long QT syndrome in Norway indicating a high prevalence of heterozygous mutation carriers. *Scand J Clin Lab Invest* 2008;**68**:362–8.

34. **Marjamaa A,** Salomaa V, Newton-Cheh C, *et al*. High prevalence of four long QT syndrome founder mutations in the Finnish population. *Ann Med* 2009;**41**:234–40.

35. **Basavarajaiah S,** Wilson M, Whyte G, *et al*. Prevalence and significance of an isolated long QT interval in elite athletes. *Eur Heart J* 2007;**28**:2944–9.

36. **Bokil NJ,** Baisden JM, Radford DJ, *et al*. Molecular genetics of long QT syndrome. *Mol Genet Metab* 2010;**101**:1–8.

37. **Schulze-Bahr E,** Haverkamp W, Borggrefe M, *et al*. Molecular genetics of arrhythmias—a new paradigm. *Z Kardiol* 2000;**89**(Suppl 4):IV12–22.

38. **Erickson CC.** Establishing risk in siblings of long QT syndrome: expressivity over penetrance. *Heart Rhythm* 2008;**5**:837–8.

39. **Deo R,** Albert CM. Epidemiology and genetics of sudden cardiac death. *Circulation* 2012;**125**:620–37.

40. **Ingles J,** Zodgekar PR, Yeates L, *et al*. Guidelines for genetic testing of inherited cardiac disorders. *Heart Lung Circ* 2011;**20**:681–7.

41. **Niimura H,** Bachinski LL, Sangwatanaroj S, *et al*. Mutations in the gene for cardiac myosin-binding protein C and late-onset familial hypertrophic cardiomyopathy. *N Engl J Med* 1998;**338**:1248–57.

42. **Quarta G,** Muir A, Pantazis A, *et al*. Familial evaluation in arrhythmogenic right ventricular cardiomyopathy: impact of genetics and revised task force criteria. *Circulation* 2011;**123**:2701–9.

43. **Bauce B,** Frigo G, Marcus FI, *et al*. Comparison of clinical features of arrhythmogenic right ventricular cardiomyopathy in men versus women. *Am J Cardiol* 2008;**102**:1252–7.

44. **Keren A,** Syrris P, McKenna WJ. Hypertrophic cardiomyopathy: the genetic determinants of clinical disease expression. *Nat Clin Pract Cardiovasc Med* 2008;**5**:158–68.

45. **Sen-Chowdhry S,** Syrris P, Pantazis A, *et al*. Mutational heterogeneity, modifier genes, and environmental influences contribute to phenotypic diversity of arrhythmogenic cardiomyopathy. *Circ Cardiovasc Genet* 2010;**3**:323–30.

46. **Watkins H,** McKenna WJ, Thierfelder L, *et al*. Mutations in the genes for cardiac troponin T and alpha-tropomyosin in hypertrophic cardiomyopathy. *N Engl J Med* 1995;**332**:1058–64.

47. **Waldmuller S,** Erdmann J, Binner P, *et al*. Novel correlations between the genotype and the phenotype of hypertrophic and dilated cardiomyopathy: results from the German Competence Network Heart Failure. *Eur J Heart Fail* 2011;**13**:1185–92.

48. **Charron P,** Dubourg O, Desnos M, *et al*. Genotype-phenotype correlations in familial hypertrophic cardiomyopathy. A comparison between mutations in the cardiac protein-C and the beta-myosin heavy chain genes. *Eur Heart J* 1998;**19**:139–45.

49. **Hwang TH,** Lee WH, Kimura A, *et al*. Early expression of a malignant phenotype of familial hypertrophic cardiomyopathy associated with a Gly716Arg myosin heavy chain mutation in a Korean family. *Am J Cardiol* 1998;**82**:1509–13.

50. **van Rijsingen IA,** Arbustini E, Elliott PM, *et al*. Risk factors for malignant ventricular arrhythmias in lamin a/c mutation carriers a European cohort study. *J Am Coll Cardiol* 2012;**59**:493–500.

51. **Sen-Chowdhry S,** Syrris P, Ward D, *et al*. Clinical and genetic characterization of families with arrhythmogenic right ventricular dysplasia/cardiomyopathy provides novel insights into patterns of disease expression. *Circulation* 2007;**115**:1710–20.

52. **Schwartz PJ,** Priori SG, Spazzolini C, *et al*. Genotype-phenotype correlation in the long-QT syndrome: gene-specific triggers for life-threatening arrhythmias. *Circulation* 2001;**103**:89–95.

53. **Vincent GM,** Schwartz PJ, Denjoy I, *et al*. High efficacy of beta-blockers in long-QT syndrome type 1: contribution of noncompliance and QT-prolonging drugs to the occurrence of beta-blocker treatment 'failures'. *Circulation* 2009;**119**:215–21.

54. **Goldenberg I,** Bradley J, Moss A, *et al*. Beta-blocker efficacy in high-risk patients with the congenital long-QT syndrome types 1 and 2: implications for patient management. *J Cardiovasc Electrophysiol* 2010;**21**:893–901.

55. **Goldenberg I,** Hall WJ, Beck CA, *et al*. Reduction of the risk of recurring heart failure events with cardiac resynchronization therapy: MADIT-CRT (Multicenter Automatic Defibrillator Implantation Trial With Cardiac Resynchronization Therapy). *J Am Coll Cardiol* 2011;**58**:729–37.

56. **Moss AJ,** Shimizu W, Wilde AA, *et al*. Clinical aspects of type-1 long-QT syndrome by location, coding type, and biophysical function of mutations involving the KCNQ1 gene. *Circulation* 2007;**115**:2481–9.

57. **Richard P,** Isnard R, Carrier L, *et al*. Double heterozygosity for mutations in the beta-myosin heavy chain and in the cardiac myosin binding protein C genes in a family with hypertrophic cardiomyopathy. *J Med Genet* 1999;**36**:542–5.

58. **Ingles J,** Doolan A, Chiu C, *et al*. Compound and double mutations in patients with hypertrophic cardiomyopathy: implications for genetic testing and counselling. *J Med Genet* 2005;**42**:e59.

59. **Bauce B,** Nava A, Beffagna G, *et al*. Multiple mutations in desmosomal proteins encoding genes in arrhythmogenic right ventricular cardiomyopathy/dysplasia. *Heart Rhythm* 2010;**7**:22–9.

60. **Marian AJ.** Modifier genes for hypertrophic cardiomyopathy. *Curr Opin Cardiol* 2002;**17**:242–52.

61. **Tomas M,** Napolitano C, De Giuli L, *et al*. Polymorphisms in the NOS1AP gene modulate QT interval duration and risk of arrhythmias in the long QT syndrome. *J Am Coll Cardiol* 2010;**55**:2745–52.

62. **Hershberger RE,** Cowan J, Morales A, *et al*. Progress with genetic cardiomyopathies: screening, counseling, and testing in dilated, hypertrophic, and arrhythmogenic right ventricular dysplasia/cardiomyopathy. *Circ Heart Fail* 2009;**2**:253–61.

63. **Ackerman MJ,** Priori SG, Willems S, *et al*. HRS/EHRA expert consensus statement on the state of genetic testing for the channelopathies and cardiomyopathies this document was developed as a partnership between the Heart Rhythm Society (HRS) and the European Heart Rhythm Association (EHRA). *Heart Rhythm* 2011;**8**:1308–39.

64. **Voelkerding KV,** Dames S, Durtschi JD. Next generation sequencing for clinical diagnostics-principles and application to targeted resequencing for hypertrophic cardiomyopathy: a paper from the 2009 William Beaumont Hospital Symposium on Molecular Pathology. *J Mol Diagn* 2010;**12**:539–51.

65. **Meder B,** Haas J, Keller A, *et al*. Targeted next-generation sequencing for the molecular genetic diagnostics of cardiomyopathies. *Circ Cardiovasc Genet* 2011;**4**:110–22.

66. **Kapplinger JD,** Landstrom AP, Salisbury BA, *et al*. Distinguishing arrhythmogenic right ventricular cardiomyopathy/dysplasia-associated mutations from background genetic noise. *J Am Coll Cardiol* 2011;**57**:2317–27.

67. **Maron BJ,** Chaitman BR, Ackerman MJ, *et al*. Recommendations for physical activity and recreational sports participation for young patients with genetic cardiovascular diseases. *Circulation* 2004;**109**:2807–16.

68. **Pelliccia A,** Zipes DP, Maron BJ. Bethesda Conference #36 and the European Society of Cardiology Consensus Recommendations revisited a comparison of US and European criteria for eligibility and disqualification of competitive athletes with cardiovascular abnormalities. *J Am Coll Cardiol* 2008;**52**:1990–6.

69. **Borjesson M,** Serratosa L, Carre F, *et al*. Consensus document regarding cardiovascular safety at sports arenas: position stand from the European Association of Cardiovascular Prevention and Rehabilitation (EACPR), section of Sports Cardiology. *Eur Heart J* 2011;**32**:2119–24.

70. **Corrado D,** Basso C, Pilichou K, *et al*. Molecular biology and clinical management of arrhythmogenic right ventricular cardiomyopathy/dysplasia. *Heart* 2011;**97**:530–9.

71. **Varnava A,** Baboonian C, Davison F, *et al*. A new mutation of the cardiac troponin T gene causing familial hypertrophic cardiomyopathy without left ventricular hypertrophy. *Heart* 1999;**82**:621–4.

72. **Christiaans I,** Lekanne dit Deprez RH, van Langen IM, *et al*. Ventricular fibrillation in MYH7-related hypertrophic cardiomyopathy before onset of ventricular hypertrophy. *Heart Rhythm* 2009;**6**:1366–9.

73. **McKenna WJ,** Stewart JT, Nihoyannopoulos P, *et al*. Hypertrophic cardiomyopathy without hypertrophy: two families with myocardial disarray in the absence of increased myocardial mass. *Br Heart J* 1990;**63**:287–90.

74. **Christiaans I,** Birnie E, Bonsel GJ, *et al*. Manifest disease, risk factors for sudden cardiac death, and cardiac events in a large nationwide cohort of predictively tested hypertrophic cardiomyopathy mutation carriers: determining the best cardiological screening strategy. *Eur Heart J* 2011;**32**:1161–70.

75. **Pasquale F,** Syrris P, Kaski JP, *et al*. Long-term outcomes in hypertrophic cardiomyopathy caused by mutations in the cardiac troponin T gene. *Circ Cardiovasc Genet* 2012;**5**:10–17.

76. **Tabib A,** Loire R, Chalabreysse L, *et al*. Circumstances of death and gross and microscopic observations in a series of 200 cases of sudden death associated with arrhythmogenic right ventricular cardiomyopathy and/or dysplasia. *Circulation* 2003;**108**:3000–5.

77. **Bauce B,** Rampazzo A, Basso C, *et al*. Clinical phenotype and diagnosis of arrhythmogenic right ventricular cardiomyopathy in pediatric patients carrying desmosomal gene mutations. *Heart Rhythm* 2011;**8**:1686–95.

78. **Bauce B,** Basso C, Rampazzo A, *et al*. Clinical profile of four families with arrhythmogenic right ventricular cardiomyopathy caused by dominant desmoplakin mutations. *Eur Heart J* 2005;**26**:1666–75.

79. **van Berlo JH,** de Voogt WG, van der Kooi AJ, *et al*. Meta-analysis of clinical characteristics of 299 carriers of LMNA gene mutations: do lamin A/C mutations portend a high risk of sudden death? *J Mol Med (Berl)* 2005;**83**:79–83.

80. **Meune C,** Van Berlo JH, Anselme F, *et al*. Primary prevention of sudden death in patients with lamin A/C gene mutations. *N Engl J Med* 2006;**354**:209–10.

81. **Ehlermann P,** Lehrke S, Papavassiliu T, *et al*. Sudden cardiac death in a patient with lamin A/C mutation in the absence of dilated cardiomyopathy or conduction disease. *Clin Res Cardiol* 2011;**100**:547–51.

82. **Pasotti M,** Klersy C, Pilotto A, *et al*. Long-term outcome and risk stratification in dilated cardiolaminopathies. *J Am Coll Cardiol* 2008;**52**:1250–60.

83. **Kirchhof P,** Fabritz L, Zwiener M, *et al*. Age- and training-dependent development of arrhythmogenic right ventricular cardiomyopathy in heterozygous plakoglobin-deficient mice. *Circulation* 2006;**114**:1799–806.

84. **Maron BJ,** Araújo CG, Thompson PD, et al. Recommendations for preparticipation screening and the assessment of cardiovascular disease in masters athletes: an advisory for healthcare professionals from the working groups of the World Heart Federation, the International Federation of Sports Medicine, and the American Heart Association Committee on Exercise, Cardiac Rehabilitation, and Prevention. Circulation 2001;**103**:327–34.

85. **Corrado D,** Pelliccia A, Björnstad HH, et al. Cardiovascular pre-participation screening of young competitive athletes for prevention of sudden death: proposal for a common European protocol. Consensus Statement of the Study Group of Sport Cardiology of the Working Group of Cardiac Rehabilitation and Exercise Physiology and the Working Group of Myocardial and Pericardial Diseases of the European Society of Cardiology. Eur Heart J 2005;**26**:516–24.

86. **Pelliccia A,** Fagard R, Björnstad HH, et al. Recommendations for competitive sports participation in athletes with cardiovascular disease: a consensus document from the Study Group of Sports Cardiology of the Working Group of Cardiac Rehabilitation and Exercise Physiology and the Working Group of Myocardial and Pericardial Diseases of the European Society of Cardiology. Eur Heart J 2005;**26**:1422–45.

87. **Zipes DP,** Ackerman MJ, Estes NA, et al. Task force 7: arrhythmias. J Am Coll Cardiol 2005;**45**:1354–63.

88. **Maron BJ,** Zipes DP. Introduction: eligibility recommendations for competitive athletes with cardiovascular abnormalities-general considerations. J Am Coll Cardiol 2005;**45**:1318–21.

89. **Pelliccia A,** Corrado D, Björnstad HH, et al. Recommendations for participation in competitive sport and leisure-time physical activity in individuals with cardiomyopathies, myocarditis and pericarditis. Eur J Cardiovasc Prev Rehabil 2006;**13**:876–85.

90. **Corrado D,** Basso C, Pavei A, et al. Trends in sudden cardiovascular death in young competitive athletes after implementation of a preparticipation screening program. JAMA 2006;**296**:1593–601.

91. **Mahon NG,** Murphy RT, MacRae CA, et al. Echocardiographic evaluation in asymptomatic relatives of patients with dilated cardiomyopathy reveals preclinical disease. Ann Intern Med 2005;**143**:108–15.