Effect of a high carbohydrate diet on core temperature during prolonged exercise

M.P. Schwellnus MBCh, MSc1, N.F. Gordon MBCh, PhD2, G.G. van Zyl BA (Hons)3, J.F. Cilliers, PhD3, H.C. Grobler, BA (Hons)3, J. Kuyl MBChB, FFPath4 and H.W. Kohl MSPH2

1 MRC/UCT Bioenergetics of Exercise Research Unit, University of Cape Town Medical School, Cape Town, South Africa
2 Institute for Aerobics Research, Dallas, Texas; Department of Chemical Pathology, University of the Witwatersrand, Johannesburg, South Africa
3 Biokinetic Centre, 1 Military Hospital, Pretoria, South Africa
4 South African Institute for Medical Research, Johannesburg, South Africa

This study compared the effects of a high-carbohydrate and a mixed diet on core temperature responses to prolonged exercise in six male competitive cyclists (age = 22.2 ± 1.9 years). This study, the first to investigate the effect of a high-carbohydrate diet on core temperature in humans, therefore suggests that three days of increased dietary carbohydrate intakes do not evoke any deleterious thermoregulatory responses during prolonged submaximal exercise.

Keywords: Exercise, diet, carbohydrates, temperature

Introduction

The practice of consuming a high-carbohydrate diet in the days preceding competitive endurance events is common amongst athletes1,2. The resultant increase in muscle glycogen stores has been shown to delay the onset of fatigue and increase the ability to perform prolonged exercise1,3,4. However, competitive endurance events are associated with an accentuated risk of heat injury, particularly when conducted in hot and humid environments5, and the effect of a high-carbohydrate diet on exercise thermoregulation has yet to be studied in humans. This is of concern because animal studies suggest that although glucose infusion during exercise may reduce thermal stress6,7, a high-carbohydrate diet might in fact exaggerate the core temperature response8. Accordingly, we investigated the effect of a high-carbohydrate diet on the thermoregulatory responses of six competitive cyclists during 150 minutes of submaximal exercise.

Materials and methods

Subjects

Six healthy male competitive endurance cyclists served as subjects. All were volunteers and gave written, informed consent. Physical characteristics of the subjects are documented in Table 1.

Maximal oxygen uptake

Each subject’s maximal oxygen uptake was determined prior to the thermoregulatory investigations. Maximal graded exercise testing was performed using a calibrated Monark cycle ergometer. The starting workload (80 W) was increased by 40 W every three minutes until volitional exhaustion. The pedal frequency was maintained at 80 r.p.m. throughout each test. During exercise, subjects breathed through a low-resistance Hans Rudolph valve and the relevant respiratory variables were determined over each minute using automated open circuit spirometry (Gould 9000 IV Computerized Pulmonary Lab). The system’s dry rolling seal spirometer, paramagnetic oxygen analyser and infra-red absorption carbon dioxide analyser were calibrated immediately before each test using standard procedures. The peak oxygen uptake attained during graded exercise testing was taken as the maximal oxygen uptake.

Dietary intervention

Each subject performed two 150 minute submaximal exercise tests on a calibrated Monark cycle ergometer, at least 10 days apart. Four days before each test,

Table 1. Physical characteristics of subjects

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>Value</th>
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<tr>
<td>Age (yr)</td>
<td>22.2 ± 1.9</td>
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<tr>
<td>Weight (kg)</td>
<td>72.46 ± 4.75</td>
</tr>
<tr>
<td>Height (cm)</td>
<td>178 ± 6</td>
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<tr>
<td>VO2 max (ml/kg/min)</td>
<td>64.0 ± 5.8</td>
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</tbody>
</table>

Values are mean ± SD; n = 6; VO2 max = maximal oxygen uptake
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Subjects completed a compulsory intense training session of at least 2 hours duration, in order to deplete their muscle glycogen stores. During the three days prior to both submaximal exercise tests subjects adhered to a specific diet, prescribed in a randomized crossover fashion. The prescribed diets were either a high-carbohydrate or a mixed diet, containing >500 g carbohydrate/day or <200 g carbohydrate/day, respectively. No strenuous physical activity was performed during the dietary intervention period.

Thermoregulatory investigation

Subjects reported to the laboratory at the same time of day on each of the two experimental days. Meals and fluids, with the exception of water, were provided for 2 hours before exercise. Water was consumed ad libitum until ±15 minutes before exercise, at which time all subjects were required to drink 200 ml of tap water. Subjects were acclimated for 60 minutes before exercise in the environmentally controlled laboratory, where the dry bulb temperature (±2 °C) and relative humidity (±55 per cent) remained constant for both tests.

Each submaximal test consisted of 150 minutes cycling, interrupted for five minutes after 60 and 120 minutes in order to facilitate nude body-weight and rectal temperature measurement. The pedal frequency was maintained at 80 r.p.m. during cycling and workloads were chosen to elicit ±50 per cent of the predetermined maximal oxygen uptake. Individual workloads were identical for both 150 minutes exercise bouts. During exercise, only cycling shorts were worn, and 200 ml of tap water was consumed every 20 minutes.

Heart rates, rectal temperature, oesophageal temperatures and nude body-weights were measured immediately before exercise and at 60, 120, and 150 minutes of exercise. During exercise tests, heart rates and oesophageal temperatures were recorded while the subjects were cycling, whereas rectal temperatures and nude body-weights were recorded after three minutes of standing rest. Heart rates were determined by counting the arterial pulse for 30 seconds. Rectal temperature was measured ±10 cm beyond the external anal sphincter. Oesophageal temperature was measured at heart level using the electrocardiographic method. Copper-constantan thermocouples were used for all temperature measurements. On completion of the study thermocouples were calibrated in a stirred water bath against a certified thermometer. Nude body-weights were recorded using a balance scale sensitive to a change of 50 g. Sweat rates were calculated from weight differences, taking water ingested and urine voided into account.

Venous blood samples were taken from an antecubital vein immediately before exercise and, while the subjects were cycling, at 60 and 150 minutes of exercise. Percentage changes in plasma volume were calculated from haemoglobin and haematocrit values using the formula of Dill and Costill. Ventilatory responses were measured over a five minute period, after 55 and 145 minutes of exercise.

<table>
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<tr>
<th>Table 2. Effect of a high-carbohydrate (C) and mixed (M) diet on cardiorespiratory responses during 150 minutes of cycling</th>
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<tr>
<td></td>
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<tr>
<td>C diet</td>
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<tr>
<td>M diet</td>
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<tr>
<td>VO₂ max</td>
</tr>
<tr>
<td>C diet</td>
</tr>
<tr>
<td>M diet</td>
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<tr>
<td>Values are mean ±SD; n = 6; HR = heart rate; RPE = rate of perceived exertion; RER = respiratory exchange ratio; VE = pulmonary ventilation; VO₂ = oxygen uptake; %VO₂ max = percentage of maximal oxygen uptake.</td>
</tr>
</tbody>
</table>

as earlier outlined. Each participant’s subjective perception of effort was evaluated, using the Borg 6–20 perceived exertion scale, at 60, 120 and 150 minutes of exercise.

Statistics

Standard analysis of variance techniques were used to assess differences between treatments and among time periods. Polynomial contrasts were used to assess the response pattern in analyses with more than two time periods. Statistical significance was established at the 0.05 probability level.

Results

As expected, nude body-weights were significantly higher (P < 0.05) with the high-carbohydrate diet (72.46 ± 4.75 kg) than with the mixed diet (71.41 ± 5.06 kg). This increase in weight most likely reflects an increase in glycogen and water storage with the high-carbohydrate diet.

The effect of dietary intervention on cardiorespiratory responses to exercise is documented in Table 2. Oxygen uptakes, whether expressed on an absolute or relative basis, were not significantly altered by the ingestion of a high-carbohydrate diet. In contrast, respiratory exchange ratios were significantly higher (P < 0.05) with the high-carbohydrate diet than with the mixed diet. The very high values obtained in this study (0.93–1.01) are probably related to the carbohydrate meal taken two hours before the exercise bout. Other possibilities are hyperventilation and poorly calibrated equipment. The latter is unlikely as
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the equipment was calibrated before and after each test. The difference in RER that was observed is indicative of an increased reliance on carbohydrate metabolism for energy production during exercise with the high carbohydrate diet.

Rectal (Figure 1) and oesophageal (Figure 2) temperature responses to exercise were essentially equivalent with the two diets. Similarly, sweat rates (Figure 3) and percentage changes in plasma volume (Table 3) were similar with the high-carbohydrate and mixed diets.

Discussion

A direct relationship between dietary carbohydrate content and the relative contribution of different metabolic substrates to energy production during exercise is well established. Recent studies have further indicated that changes in fuel utilization during exercise may modify thermoregulatory responses. More specifically, glucose infusion during prolonged exercise has been shown to attenuate core temperature rises in several animal studies. In contrast, exaggerated core temperature responses to exercise have been observed in animals as a result in insulin-induced hypoglycaemia or inhibition of glucose utilization by 2-deoxy-D-glucose administration. Although these animal studies suggest that an enhanced contribution of carbohydrates to muscle metabolism may reduce exercise hypothermia, Owen et al. were unable to demonstrate a significant effect of carbohydrate ingestion during exercise on thermoregulation in humans. Moreover, Francesconi and Hubbard have documented findings that are in fact contrary to the hypothesis that the rise in core temperature during exercise can be attenuated by an increased contribution of carbohydrates to muscle metabolism. In their study, rats placed on a high-carbohydrate diet for four days prior to exercise in heat displayed higher resting and exercise core temperatures than those on a normal diet.

When extrapolated to humans, the results of the study conducted by Francesconi and Hubbard imply that the practice of consuming a diet high in carbohydrates during the days preceding competitive endurance events may actually accentuate the risk for hyperthermia. However, in our study, the first to investigate the effect of a high-carbohydrate diet on exercise thermoregulation in humans, three days of increased dietary carbohydrate intake failed to

### Table 3. Effect of a high-carbohydrate and mixed diet on the percentage change in plasma volume during 150 minutes of cycling

<table>
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<tr>
<th></th>
<th>% Change in Plasma Volume</th>
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<tbody>
<tr>
<td></td>
<td>0–60 min</td>
</tr>
<tr>
<td>High-carbohydrate</td>
<td>-2.6 ± 6.1</td>
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<tr>
<td>Mixed</td>
<td>-2.8 ± 3.9</td>
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Values are mean ± SD; n = 6

Values did not differ significantly
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significantly alter rectal and oesophageal temperatures, sweating, or plasma volume changes during 150 minutes of submaximal cycling performed in a cool environment. On the basis of our data, it therefore appears that an increased contribution of carbohydrates to muscle metabolism as a result of a high-carbohydrate diet does not evoke any deleterious thermoregulatory consequences during prolonged exercise in humans. However, additional research involving more intense exercise and less favourable environmental conditions will be needed to fully clarify the situation.

References