Carbohydrate loading in practice: high muscle glycogen concentration is not certain

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It is believed that muscle glycogen resynthesis can be stimulated by depleting the glycogen stores by heavy physical exercise and then eating a diet rich in carbohydrates. In this study, we compared muscle glycogen concentrations after two different depletion and loading procedures in six male runners. The depletion runs for the procedures were a half-marathon race and an easier fartlek. The mean muscle glycogen concentrations (±s.e.m.), analysed after the procedures, did not differ significantly between the race and the fartlek being 285 (±25) mmol/kg d.w. (dry weight) versus 315 (±32) mmol/kg d.w. (P > 0.05). Moreover, the subjects’ glycogen concentrations were not clearly increased above the prederepletion values following either procedure. The results show that higher glycogen levels do not necessarily occur after classical carbohydrate-loading procedures.

Keywords: Carbohydrate diet, hormones, marathon

Studies in the late 1960s showed a positive correlation between the pre-exercise muscle glycogen concentration and the ability to perform prolonged, severe exercises. An elevation of muscle glycogen stores can be achieved by a manipulation of the diet. In the classical method, muscle glycogen stores are first depleted by prolonged, exhaustive exercise, followed by 2–3 days of low-carbohydrate diet, i.e. fat and protein mainly. Then the stores are replenished by eating a high-carbohydrate diet and taking only light exercise.

The exhaustive depletion exercise has been criticized, because it interferes with the peaking for an important race. Sherman and Costill and Blom et al. have shown that an exhaustive depletion brought about by running might even be totally ineffective in stimulating glycogen synthesis. Despite this, the traditional procedure (exhaustive depletion followed by a carbohydrate-rich diet) is carried out by many endurance athletes as a final preparation for a race.

The aim of the present study was to find out whether muscle glycogen concentrations would differ after two dissimilar carbohydrate-loading procedures, performed in field conditions. The depletion exercises for the procedures involved: a half-marathon race and an easier fartlek. Both procedures are commonly used by marathon runners.

We also investigated the extent of the physical strain caused by the two depletion runs. Changes in serum testosterone, free testosterone and cortisol concentrations are used as indicators of strain. The rationale for measuring these hormones is that serum testosterone and free testosterone concentrations have been shown to decrease and serum cortisol concentrations to increase as a consequence of heavy physical exercise.

Subjects and methods

Subjects and experimental design

After being informed of the experimental procedures and inherent risks, six well trained, national level, endurance runners (age 29 (±2) years, weight 66 (±2) kg, height 171 (±1) cm and training 552 (±35) km during the previous month – all figures being mean (±s.e.m.)) gave their written consent to participate in the study. Easy training, no more than 1 h per day, was allowed for the 3 days before the depletion run. The subjects performed two carbohydrate-loading procedures, but the depletion runs were different. Twenty-one days separated the two depletion runs, in order to avoid possible carry-over effects from the first procedure.

Depletion and loading

In the ‘race’ procedure, the depletion run involved a half-marathon road-race on relatively flat terrain. The subjects ran the distance in 74.2 (±0.5) min and their running speed was 3.5 min/km (4.8 m/s), roughly equivalent to 70–80% of their \( V_{O_2\ max} \). The race was followed by maximal 200 m sprints until voluntary exhaustion occurred (about 15 sprints). Their total running time was 95–100 min. In the ‘fartlek’ procedure the running course differed and it was not totally flat; the athletes’ total running time was the same as for the race (95–100 min), but their mean...
Carbohydrate loading in practice: G. M. Fogelholm et al.

Table 1. Experimental procedures

<table>
<thead>
<tr>
<th>Procedure 1: Race</th>
<th>Procedure 2: Fartlek</th>
<th>Samples</th>
</tr>
</thead>
<tbody>
<tr>
<td>Day—1 Easy running 60 min</td>
<td>Easy running 60 min</td>
<td></td>
</tr>
<tr>
<td>Depletion Day 0</td>
<td>Half-marathon race (3.5 min/km and 10–20 x 200 m until exhaustion)</td>
<td>100 min running (4.0 min/km)</td>
</tr>
<tr>
<td>Day 1</td>
<td>Running 80–90 min</td>
<td>Running 80–90 min</td>
</tr>
</tbody>
</table>

Ba, post-run blood samples taken in the afternoon; Bm, fasting blood sample taken in the morning; Fr, food record; M, muscle biopsy taken in the morning.

Running speed was slower (4.0 min/km or 4.2 m/s), close to their normal training pace.

On the first day after the depletion run, an 80–90 min run was executed. The subjects ate a reduced carbohydrate diet for these 2 days. During the 4-day carbohydrate-loading phase, the subjects executed one easy 45–60 min (4.5 min/km) run per day. The experimental procedures are shown in Table 1.

The athletes were instructed on how to choose foods and liquids that would ensure a reduced-carbohydrate diet during the depletion phase. More detailed instructions were provided for the high-carbohydrate diet. All subjects were instructed how to compose a basic diet that would ensure a daily carbohydrate intake of 400 g. They were then able to choose from several different food items, providing additional carbohydrate. This was to make certain that they increased their daily intake of carbohydrate to 9 g/kg body weight. The daily nutrient intake was calculated from the subjects’ food diaries by a computer program. To aid their food-recording process, all participants were given postal scales for weighing portions. During loading, there were no significant differences in the carbohydrate intake between the intense and moderate procedures (Table 2). The mean percentages of carbohydrate in the total energy intake were 70% for the race procedure and 67% for the fartlek.

Table 2. Energy and carbohydrate intake of six marathon runners during two carbohydrate-loading procedures. Results are expressed as mean (± s.e.m.).

<table>
<thead>
<tr>
<th>Procedure 1: Race</th>
<th>Procedure 2: Fartlek</th>
</tr>
</thead>
<tbody>
<tr>
<td>Energy (MJ/day)</td>
<td></td>
</tr>
<tr>
<td>Depletion (days 0–1)</td>
<td>9.2 (±1.1)</td>
</tr>
<tr>
<td>Loading (days 2–5)</td>
<td>14.3 (±0.8)</td>
</tr>
<tr>
<td>Carbohydrates (g/day)</td>
<td></td>
</tr>
<tr>
<td>Depletion (days 0–1)</td>
<td>205 (±68)</td>
</tr>
<tr>
<td>Loading (days 2–5)</td>
<td>598 (±44)</td>
</tr>
</tbody>
</table>

* Significant difference (P < 0.05) between the procedures

Blood analyses

All blood samples were taken after a 15 min rest. The venous blood samples were drawn from the antecubital vein with the subjects in a sitting position. After an overnight fast, the morning samples were taken between 8.00 and 10.00 hours. Samples were also drawn within 1 h after each depletion run. Only water, in unlimited amounts, was allowed between the run and the time of taking the sample.

Radioimmunological methods were used for the measurement of serum cortisol and testosterone. Serum sex hormone-binding globulin (SHBG) concentrations were determined by an immunoradiometric (IRMAs) method (Farmos Diagnostica, Oulunsalo, Finland). Serum free-testosterone concentrations were calculated using testosterone and SHBG values. All measurements were carried out in duplicate, and in order to avoid interassay variation, all the assays of each subject were run in the same series. The intra-assay coefficients of variation for the analyses were 9.1% for plasma cortisol, 8.3% for plasma testosterone and 8.8% for SHBG.

Muscle analyses

After local anaesthesia of the skin with lignocaine, without adrenaline, muscle biopsies were taken from the lateral portion of the quadriceps femoris muscle. Because we had previously taken several consecutive biopsies from that muscle, without the subjects experiencing complications, the vastus lateralis was chosen as the biopsy site. The muscle samples were taken before the depletion runs and at the end of the loading phase. The concentration of muscle glycogen was measured from 50–350 µg of freeze dried cryostat sections after alkaline digestion and ethanol precipitation. The samples were then analysed for glucose.

Statistical analyses

All results are expressed as mean (± s.e.m.). Analysis of variance for repeated measurements (BMDP 2V-package) was used for the statistical analyses. Significant (P < 0.05) time, procedure or time × procedure effects were further identified by Wilcoxon’s signed rank test.

Results

Normal muscle glycogen concentrations were observed before both procedures: the group mean values were 343 (±10) mmol/kg d.w. before the race procedure, and 345 (±11) mmol/kg d.w. before the fartlek. There were no differences (P > 0.05) in glycogen concentrations resulting from the two carbohydrate-loading procedures (Figure 1): the range of post-loading muscle glycogen concentration was 170–350 mmol/kg d.w. after the race, and 217–393 mmol/kg d.w. after the fartlek. The two very low glycogen levels (one after the race, the other after the fartlek) were not found in the same subject. After a total of 12 depletion and loading procedures (two
carbohydrate loading in practice: G. M. Fogelholm et al.

Figure 2. Serum free-testosterone concentration during two glycogen depletion and loading procedures in six male runners. The procedures differed in the depletion run which was either hard (race) or moderate (fartlek). The results are expressed as mean ± s.e.m. Significant differences (P < 0.05) between the procedures and the changes from pre-run values are denoted with an asterisk. ■, Moderate run; □, hard run

Discussion

In the present study, neither the race nor the fartlek procedure resulted in clearly increased glycogen concentrations in the subjects. In addition, no differences in final muscle glycogen levels between

for each subject) were carried out, only five resulted in increased glycogen levels.

A significant (P < 0.05) decrease in the serum free-testosterone concentrations (Figure 2) was observed after both depletion runs, but the decrease was more pronounced (P < 0.05) after the race run. Even 6 days after the half-marathon race, at the end of the loading phase, the free-testosterone concentrations were still significantly lower than the pre-run values. In five out of six subjects, the serum cortisol concentrations were higher after the race than those obtained after the fartlek. This difference did not reach statistical significance.

Discussion

In the present study, neither the race nor the fartlek procedure resulted in clearly increased glycogen concentrations in the subjects. In addition, no differences in final muscle glycogen levels between

the procedures were observed. The failure to demonstrate an increase in glycogen concentration following either experiment was rather surprising, since these kinds of procedures are commonly carried out by marathon runners in Finland. It seems that higher glycogen concentrations, in the vastus lateralis muscle, do not necessarily occur after carbohydrate-loading procedures in field conditions. Hence, the results agree well with the laboratory studies carried out by Blom et al.

We admit that the easy runs, performed daily by all subjects during the loading phase, may be one reason why much higher muscle glycogen concentrations did not occur. However, we did not want the subjects to rest because, in practice, marathon runners often perform easy exercises during the loading phase. A study to compare glycogen levels after complete rest and after easy running during the loading phase would be interesting.

The muscle biopsies in the present study were taken from the vastus lateralis muscle, because we were familiar with the technique. It is possible that glycogen synthesis occurs more readily in the gastrocnemius muscle. On the other hand, Karlsson and Saltin found increased glycogen levels in the vastus lateralis after a carbohydrate-loading regimen. Moreover, in the study of Blom et al., the results (normal glycogen concentrations, despite carbohydrate loading) were the same as ours, although they used biopsies taken from the gastrocnemius muscle. They also theorized that muscle fibre damage, sometimes associated with prolonged running, might interfere with glycogen resynthesis.

We doubt that the diets unduly influenced the above findings. We did not use a strict fat and protein diet during the depletion phase, but this should not

Figure 1. Individual changes of muscle glycogen concentrations in the lateral portion of the quadriceps muscle before and after two depletion and loading procedures performed by six male runners: a procedure 1, race; b procedure 2, fartlek.
Carbohydrate loading in practice: G. M. Fogelholm et al.

impede glycogen resynthesis\textsuperscript{23}. The carbohydrate intake, about 600 g/day during the fartlek and race procedures, should have been enough to ensure a maximal rate of glycogen synthesis\textsuperscript{2}. Moreover, 40% of total carbohydrate was derived from whole-grain cereals, which contain a lot of complex carbohydrates. Complex carbohydrates might even stimulate glycogen resynthesis more effectively than mono- or disaccharides\textsuperscript{3}. In other studies, a lower intake of carbohydrate has resulted in clearly increased glycogen concentrations in the vastus lateralis muscle\textsuperscript{24, 25, 27}.

All the subjects felt that the fartlek run was much less exhausting than the race. This subjective feeling of fatigue after the intense run was confirmed by hormonal analyses: serum testosterone and free testosterone concentration decreased after the intense run, which is in agreement with Dessypris et al.\textsuperscript{24} and Kuoppasalmi et al.\textsuperscript{7}. Increased serum cortisol values, observed in five out of six subjects after the intense run, further indicated heavy physical strain.

It can be conjectured that the low concentrations of free testosterone throughout the loading phase after the race impaired glycogen resynthesis. In rats, low serum testosterone concentrations may cause a reduction in glycogen synthesis\textsuperscript{30, 29}. However, it is not known whether this occurs in man as well.

In conclusion, we studied a total of 12 glycogen depletion and loading procedures in six athletes in field conditions, and only five out of 12 resulted in higher glycogen concentrations in the vastus lateralis muscle after rather than before the procedure. Different depletion runs did not affect post-loading glycogen levels. Since an exhaustive depletion exercise might lead to physical overstrain, we do not recommend it for marathon runners.

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References