Leptin may be a nutrient sensing signal of adipose and muscle tissue; it is thought to be the missing link between adipose tissue and certain systems regulating body mass and energy expenditure. Its major role is to relay information on the status of energy stores to signal transducing receptors in the hypothalamus, thus regulating eating. The greater the amount of adipose tissue present, the more leptin is produced and released into the circulation. The role of leptin in the regulation of energy homeostasis has been the focus of several investigations. Exercise is a potent stimulus for secretion of many hormones, and exercise mediated negative energy balance may contribute to the regulation of plasma leptin concentrations. Although prolonged physical activity has been reported to reduce plasma leptin concentrations, the effect of physical activity, independently of loss of body fat mass, has not yet been clearly established. However, recent studies have shown that endurance exercise with marked negative energy balance—for example, marathon swimming, marathon running—may cause an immediate or delayed decrease in leptin concentration. In contrast, short term (≤30 minutes) exercise studies suggest that leptin concentration is not acutely affected by exercise regardless of intensity, as the exercise mediated energy expenditure may not reach the critical level needed for plasma leptin concentrations to be reduced by acute exercise. In addition, these short term exercise studies have used exercise protocols in which only the lower extremities were involved—that is, running or cycling. To our knowledge, no studies have yet investigated the effect of short term maximal exercise on plasma leptin using an exercise protocol in which all major muscle groups are involved. Therefore we evaluated the effects of 30 minutes of maximal ergometer rowing on plasma leptin in male rowers. We hypothesised that maximal ergometer rowing would have an impact on plasma leptin concentrations, as rowers have relatively large body masses and all extremities and trunk muscles are involved in rowing.

MATERIALS AND METHODS

Study design

Thirteen male college level rowers volunteered. They had trained regularly for the preceding 5.8 (1.9) years (mean (SD)). Measurements were performed during the preparatory period. The participants trained 4.5 (2.9) times a week throughout the year. They were fully familiarised with the procedures before providing their written consent to participate in the experiment, as approved by the medical ethics committee of the University of Tartu. First, each subject completed body composition and performance assessment tests. The second measurement session consisted of a 30 minute maximal test on a rowing ergometer to determine leptin responses to relatively short term exercise. The two sessions were separated by at least one week. The rowers were asked not to participate in any physical activity in the 24 hours before each session.

Assessment of body composition

The height (Martin metal anthropometer) and body mass (A&D Instruments Ltd, Abingdon, Oxfordshire, UK) of the participants were measured to the nearest 0.1 cm and 0.05 kg respectively. Body composition was measured using dual energy x ray absorptiometry. Scans of the whole body were performed on each of the subjects using a Lunar DPX-IQ scanner (Lunar Corporation, Madison, Wisconsin, USA), and analysed for fat and fat free mass.

Exercise tests

Exercise tests were performed on a rowing ergometer (Concept II, Morrisville, North Carolina, USA) at the same time of day (between 1100 and 1300), and the time of the test was identical for each subject across both tests. All subjects had eaten a meal about two hours before the test.

Abbreviations: IGF-I, insulin-like growth factor-I; IGFBP-3, IGF-binding protein-3
the rowing performance test, the rowers were asked to cover a distance of 2000 m in the least time possible.\(^6\) The second exercise test was a 30 minute maximal rowing ergometer test, and the distance covered was considered to be the total work performed.\(^7\) Heart rates were recorded during both exercise tests using Sporttester Polar Vantage NV (Kempele, Finland).\(^8\,\)\(^9\)

### Blood analysis

A 10 ml blood sample was obtained from an antecubital vein, with the subject in the upright position, before, immediately after, and 30 minutes after the maximal rowing ergometer test.\(^7\) Similarly to other recent studies, no control trial was conducted, as diurnal changes in measured hormones were considered not to occur during this short time period.\(^2\)\(^,\)\(^3\)\(^,\)\(^4\)\(^,\)\(^14\) The plasma was separated and frozen at \(-20^\circ\)C for later analysis. Leptin was determined in duplicate by radioimmunoassay (Milenia Biotec GmbH, Bad Nauheim, Germany). This assay has a detection limit of 0.01 ng/ml, and intra- and interassay and interassay coefficients of variation of less than 5% and less than 7.5%, respectively.\(^6\)\(^9\)\(^,\)\(^10\) Insulin, growth hormone, insulin-like growth factor-I (IGF-I), and IGF-binding protein-3 (IGFBP-3) were determined in duplicate on Immulite 2000 (DPC, Los Angeles, California, USA). The intra-assay and interassay coefficients of variation for insulin were 4.5% and 12.2% respectively at an insulin concentration of 6.6 μIU/ml. The intra-assay and interassay coefficients of variation for growth hormone, IGF-I, and IGFBP-3 were less than 7%. All samples were run on the same assay. Glucose was measured by the hexokinase/glucose-6-phosphate-dehydrogenase method with a commercial kit (Boehringer, Mannheim, Germany). The intra-assay and interassay coefficients of variation for glucose were 5.3% and 12.5% respectively.\(^11\)\(^,\)\(^12\) Aliquots of whole blood were also analysed in quadruplicate for packed cell volume at 12 000 rpm for five minutes and for haemoglobin using a Lange microanalyser. Changes in plasma volume after exercise were calculated using the formulae of Dill and Costill.\(^14\)

### Statistical analysis

Means (SD) were determined. Friedman analyses of variance by ranks were used to examine changes, as the row data and their logarithmic transformations were not normally distributed. The Wilcoxon matched pairs signed ranks test was used where post hoc analysis was relevant. Regression analyses, Spearman correlation, and partial correlation coefficients were used to evaluate associations among different variables. Statistical significance was accepted at p<0.05.

### RESULTS

Table 1 presents the subject characteristics. The 2000 m rowing ergometer performance time (406.5 (15.8) seconds) was found to correspond to a mean heart rate of 184 (7) beats/min. The distance covered during the 30 minute maximal rowing ergometer test was 7870.4 (443.3) m with a mean heart rate of 175 (9) beats/min or 95.1 (2.2)% of the maximal rowing ergometer test. Blood lactate concentration before the 30 minute maximal rowing ergometer test was 1.8 (0.4) mmol/l, which was significantly increased immediately after the test (14.9 (4.3) mmol/l). It remained significantly raised after the first 30 minutes of recovery (4.8 (1.3) mmol/l). Plasma volume was reduced immediately after the ergometer test (−4.7 (5.4%); p<0.05). After 30 minutes of recovery, it had almost returned to baseline values (−0.1 (4.9)%).

The concentration of leptin and growth hormone were significantly decreased and increased respectively immediately after the maximal 30 minutes of ergometer rowing (table 2). Leptin concentration remained significantly reduced after 30 minutes of recovery. No significant changes were noted in other blood variables measured immediately after the test and after the first 30 minutes of recovery.

Regression analysis showed a positive relation between significantly decreased plasma leptin concentration immediately after the 30 minutes of maximal ergometer rowing and the distance covered (R\(^2\) = 0.645; p<0.05). In addition, changes in plasma leptin concentrations were related to changes in IGF-I concentrations immediately after the test (R\(^2\) = 0.390; p<0.05). There were no correlations between plasma leptin and other measured blood variables immediately after the test, nor were changes in leptin related to changes in other measured blood biochemical values after the test.

According to the regression analysis, percentage body fat explained 89.6% (R\(^2\) × 100; p<0.05) of the variance in basal leptin concentration. Basal leptin correlated significantly only with resting body mass (r = 0.64) and plasma insulin (r = 0.88). After normalisation for body fat, the relation between leptin and body mass was no longer significant (table 3). However, the relation between leptin and insulin remained positively correlated. In addition, basal leptin concentration was related to the training history after normalisation for body fat.

### DISCUSSION

The main purpose of the present study was to find possible effects of 30 minutes of acute exercise\(^3\)\(^,\)\(^4\) on leptin concentration in college level male rowers. Most previous studies on the effects of short term exercise on leptin have shown no change in leptin concentration regardless of exercise intensity in healthy men and women.\(^2\)\(^,\)\(^4\) Furthermore, Kraemer et al\(^6\) argued that the absence of any reduction in leptin found in short term exercise studies may be due to the limited energy expenditure during these exercise bouts and/or the exercise protocol. However, our study was designed to overcome these limitations in that the subjects were rowers, who have relatively large body masses, and all extremities and trunk muscles are involved in rowing.\(^7\)\(^,\)\(^8\) In contrast with this study, previous studies have used exercise protocols

<table>
<thead>
<tr>
<th>Variable</th>
<th>Before</th>
<th>After</th>
<th>After 30 min</th>
</tr>
</thead>
<tbody>
<tr>
<td>Leptin (ng/ml)</td>
<td>2.7 (0.6)</td>
<td>2.1 (0.8)</td>
<td>2.0 (0.9)</td>
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<tr>
<td>Growth hormone (μIU/l)</td>
<td>0.6 (0.9)</td>
<td>4.6 (5.4)</td>
<td>1.9 (2.7)</td>
</tr>
<tr>
<td>Insulin (μIU/l)</td>
<td>8.8 (6.9)</td>
<td>6.8 (2.7)</td>
<td>6.2 (4.1)</td>
</tr>
<tr>
<td>IGF-I (mg/l)</td>
<td>232.6 (100.0)</td>
<td>251.1 (59.0)</td>
<td>181.7 (157.6)</td>
</tr>
<tr>
<td>IGFBP-3 (mg/l)</td>
<td>4281.4 (766.1)</td>
<td>4482.4 (402.2)</td>
<td>3961.5 (1039.3)</td>
</tr>
<tr>
<td>Glucose (mmol/l)</td>
<td>4.5 (0.5)</td>
<td>4.9 (0.6)</td>
<td>4.4 (0.6)</td>
</tr>
</tbody>
</table>

Values are mean (SD). *Significantly different from before exercise; p<0.05.


Table 1 presents the subject characteristics. The 2000 m rowing ergometer performance time (406.5 (15.8) seconds) was found to correspond to a mean heart rate of 184 (7) beats/min.
in which only lower extremities were involved—that is, running or cycling. To our knowledge, this is the first study to investigate the effects of short term exercise on leptin concentration using a design in which the major muscle groups of the whole body are working. Accordingly, the main finding of this study was that leptin was significantly decreased immediately after the 30 minutes of maximal rowing ergometer exercise in college level male rowers. Furthermore, the distance covered was significantly related to leptin concentration immediately after the exercise.

The distance covered explained 64.5% (p<0.05) of the variance (R² = 0.645) in post-exercise plasma leptin concentration. This shows that the total work output during this exercise session was above the threshold reduction in energy availability that must be reached to alter the leptin concentration. Indeed, the blood lactate concentration immediately after 30 minutes of maximal ergometer rowing was very high (14.9 (4.3) mmol/l) and remained significantly raised for the first 30 minutes of recovery (4.8 (1.3) mmol/l), indicating high metabolic effort. In addition, the raised blood lactate concentrations show that the 30 minutes of rowing was highly anaerobic. It is known that anaerobic activity results in greater energy expenditure as a result of acute exercise. In contrast, Weltman et al showed that acute treadmill exercise for 30 minutes at, above, and below the lactate threshold did not alter leptin concentration in young men. Thus the sustained anaerobic nature of the exercise in our study may explain the significant reductions in leptin concentration immediately after the test.

It can be speculated that the data presented here (table 2) show that the amount of muscle tissue used during a short term exercise bout can also influence the leptin response. It is well known that about 70% of whole body muscle mass is involved in rowing, and the energy expended during 30 minutes of maximal ergometer rowing was enough to cause changes in leptin concentration in sportsmen with relatively high body fat (percentage body fat 13.3 (5.8)). The results of our study show that leptin could be used as a biological marker of physical stress after acute high intensity short term exercise in rowers, who use all the major muscle groups of the body during rowing.

Plasma leptin concentrations have usually been corrected for changes in plasma volume after the exercise session, as a decrease in plasma volume could increase the value. However, it may be the concentration of the hormone at the target tissues that is of importance, regardless of how the change in concentration is established. In these studies, leptin concentrations were corrected by applying a mathematical formula for haemoconcentration. Similarly to other short term exercise studies, we made no corrections to the plasma leptin concentrations after the rowing ergometer test. There was a mean 4.7% reduction in plasma volume during the exercise session. However, during the first 30 minutes of recovery, it returned to the baseline value. This further confirmed that the significantly decreased leptin concentration after exercise observed in our study was due to alterations in leptin mass rather than changes in haemoconcentration.

Similarly to the results of other studies with different athletes, basal leptin in these male college level rowers was relatively low. An interesting finding of this study is that the training history of the subjects was significantly related to the circulating leptin concentration normalised for body fat (r = −0.60; p<0.05). This further confirms the independent effect of chronic physical activity on plasma leptin concentration. In accordance with this, it has been suggested that leptin concentrations are reduced after 10 months of endurance type training in obese men, independent of body fat.

The primary regulator of leptin (ob) gene expression and secretion is adiposity, as also shown in this study (R² = 0.896; p<0.05). However, acute and chronic changes in energy balance can disproportionately downregulate or upregulate leptin secretion respectively. It has been argued that the diurnal rhythm of leptin is not dependent on energy intake or expenditure but rather on energy and/or carbohydrate availability. Increasing evidence supports the notion that insulin may be a critical regulator of ob gene expression and is a leptin secretagogue. Thus insulin may provide a mechanism by which adipose tissue detects changes in overall energy balance, and, in turn, upregulates or down-regulates ob gene expression accordingly. Similarly to other investigations, a significant correlation between leptin and insulin (r = 0.88; p<0.05) was found in this study, and this was evident even after normalisation for body fat (r = 0.82; p<0.05). Thus the relation between basal leptin and insulin concentrations appears to be independent of adipose tissue in college level male rowers. The effect of an insulin dependent mechanism on leptin was also confirmed by stepwise regression, as we found that a post-exercise IGF-1 concentration explained 39.0% (p<0.05) of the variance (R² = 0.39) in significantly decreased leptin concentration immediately after the 30 minutes of maximal ergometer rowing. This is in accordance with the study of Elias et al who argued that the decline in plasma leptin concentrations immediately after acute exercise to exhaustion may be related to the increased IGF-I concentrations in male unconditioned volunteers. However, in contrast with previous studies, changes in growth hormone concentrations did not affect changes in leptin concentration as a
result of short term maximal ergometer rowing exercise in male college level rowers. This is in line with the findings of Weltman et al., who also suggested that the growth hormone response to acute exercise is not mediated through leptin as a metabolic signal. In addition, it has been suggested that IGF-I and IGFBP-3 responses to acute exercise seem to be unrelated to growth hormone responses. Furthermore, Sonksen argues that circulating IGF-I should be considered more as a marker of growth hormone action rather than a mechanism by which growth hormone exerts its effects. However, further studies are needed to determine the individual roles of specific hormones that influence leptin concentration as a result of physical exercise.

In summary, this study shows that circulating plasma leptin is lowered as a result of 30 minutes of maximal rowing ergometer exercise in which all major body muscles are involved. The total amount of mechanical work performed by the muscles explained 64.5% of the post-exercise plasma leptin concentration. It is suggested that plasma leptin could be used as a biological marker of physical stress after short term (<30 minutes) high intensity exercise when the total work output is above the threshold reduction in energy availability.

Authors' affiliations
J. Jürimäe, T. Jürimäe, Institute of Sport Pedagogy and Coaching Sciences, University of Tartu, Tartu, Estonia

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REFERENCES